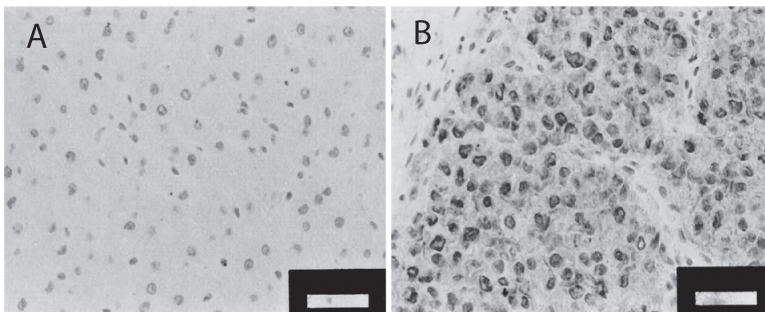


REGENERATIVE ENGINEERING FOR CANCER



Cell nucleus density and size in control rat liver and hepatoma: (A) control rat liver; (B) hepatoma. Scale bar: 50 μ m. (Reprinted with permission from la Cour JM et al: Up-regulation of ALG-2 in hepatomas and lung cancer tissue, *Am J Pathol* 163:81–9, copyright 2003.) See color insert.

CLASSIFICATION OF CANCERS [25.1]

Cancer is a disorder characterized by the formation of immortal cells that undergo uncontrolled excessive cell proliferation. Cancer cells are aberrantly differentiated cells due to gene mutation and do not possess physiological functions. These cells can aggressively invade neighboring tissues and organs, spread to remote tissues and organs through the lymphatic and vascular systems (a process known as *metastasis*), form colonies in invaded tissues and organs, and expand in the expense of normal cells. As a result, normal cells die because of deprivation of necessary nutrients, leading to the malfunction

and death of involved tissues and organs. It should be noted that cancer is a malignant type of tumor. There are also tumors that grow slowly, do not invade neighboring tissues, do not metastasize, and do not significantly influence the functions of host tissues and organs. This type of tumor is defined as benign tumors and will not be covered in this book.

Based on the origin of cancer formation, cancers are classified into several types: carcinoma, sarcoma, leukemia, and neural tumors. *Carcinoma* is a type of cancer that arises from epithelial cells, including gland cells, and can be found in a variety of organs, such as the nasal and oral cavities, gastrointestinal tract, liver, pancreas, lung, breast, prostate gland, ovary, uterus, bladder, and skin. Carcinoma is the most common type of cancer, accounting for about 80% of total cancers. The high incidence of carcinoma is possibly related to the high turnover or renewal rate of epithelial cells. These cells are developed for the protection of tissues and organs, and are often subject to various types of harmful environmental factors. Thus, mature epithelial cells are still able to differentiate and proliferate for the repair and replacement of injured and lost cells. Such a feature naturally increases the rate of cancer formation. Various terms have been used to describe carcinoma, depending on the type of cells, tissues, and organs where cancers arise. For instance, *hepatoma* is a term for liver carcinoma, *melanoma* is used to describe carcinoma derived from the epidermal melanocytes, which contains the pigment melanin, and *adenocarcinoma* is cancer from gland epithelial cells.

Sarcoma is a type of cancer that arises from connective and muscular tissues. This type of cancer is found in soft connective tissues, bones, cartilages, skeletal muscles, and blood vessels. Since mature connective tissue cells and muscular cells are well-differentiated cells and undergo a low rate of proliferation, the incidence of sarcoma (about 1%) is much lower than that of carcinoma. *Leukemia* is a type of cancer derived from hematopoietic or blood cells, primarily involving leukocytes. Leukemia derived from lymphocytes is called *lymphoma*. This type of cancer accounts for about 9% of the total cancers. Another major type of cancer is *neural cancers*, which are found in the nervous system. This type of cancer increases the volume of the brain within the limited skull space and induces the compression of normal brain tissue, resulting in various symptoms of neural disorders depending on the region involved. Typical nervous cancers include gliomas and retinoblastomas.

PATHOGENESIS OF CANCERS [25.1]

While the pathogenesis of cancers remains a research topic, increasing investigations have suggested that cancers are possibly a result of gene mutation or changes in DNA structure. Cancer may be originated from a single cell that undergoes cancerigenic gene mutation. It is important to note that, although various types of gene mutation may occur under physiological conditions, not all gene mutations lead to carcinogenesis. However, it remains poorly understood what types of gene mutation are carcinogenic.

Gene mutation may occur spontaneously during cell division as a natural process, which is responsible for evolutionary alternations in living organisms. In addition, gene mutation can be induced by environmental factors, including radiation, viral infection, exposure to carcinogens, and therapeutic gene transfer. Radiation (e.g., X rays and ultraviolet light) often causes DNA damage and chromosome disruption. Gene mutation may

be introduced to the genome when damaged genes and chromosomes are repaired. Viruses are able to integrate their genome into the host genome, a process often inducing gene mutation. Chemical carcinogens, such as formaldehyde and peanut mold-toxin (aflatoxin), can cause changes in DNA sequences. Gene transfer is thought a therapeutic method that is used to correct mutant genes. However, the insertion of foreign genes into the genome may induce gene mutation. Cancers may be induced when patients are frequently exposed to these factors.

While many types of gene mutation can contribute to carcinogenesis, there are two types that play a major role: the activation of growth stimulatory genes, which are also known as protooncogenes, and the suppression of growth inhibitory genes, which are also called tumor suppressor genes. *Protooncogenes* are normal genes that encode mitogenic proteins and can be converted to carcinogenic oncogenes or cancer-inducing genes by mutation. Examples of proto-oncogenes include c-fos (activator protein-1 gene), c-jun (another activator protein-1 gene), c-raf (protein serine/threonine kinase gene), c-myc (gene-regulatory protein gene), c-sis (platelet-derived growth factor gene), and c-src (Src protein tyrosine kinase gene). These genes play critical roles in the regulation of physiological development and morphogenesis. These genes can be transformed into oncogenes in response to the stimulation of environmental factors. A typical cause of protooncogene transformation is retrovirus infection. Retroviruses can convert their RNA genome into DNA and insert the converted DNA into selected protooncogenes in the host genome. The protooncogenes can be structurally altered or can be subject to the control of viral gene promoters, leading to the formation of carcinogenic oncogenes that stimulate cell differentiation and proliferation.

Another type of gene mutation is the alteration or loss of tumor suppressor genes. These genes play critical roles in the inhibition of carcinogenesis. The mutation or loss of such genes contributes to the initiation and development of cancers. A typical tumor suppressor gene is the retinoblastoma tumor suppressor gene. This gene is expressed in almost all cell types and encodes a protein that controls the progression of the cell division cycle. Cancers in several organs, including the lung and breast, are associated with reduced expression of the retinoblastoma tumor suppressor gene. The mutation and loss of this gene contributes to carcinogenesis. Another tumor suppressor gene is the p53 gene (see page 236 for the characteristics of p53). Patients with mutation or loss of this gene are susceptible to carcinogenesis. Some viruses, such as papillomaviruses, exert an inhibitory effect on the tumor suppressor genes.

PATHOLOGICAL CHARACTERISTICS OF CANCERS [25.1]

In pathological examinations, cancerous changes can be found at the cellular and tissue levels. Pathological changes at the cellular level include an increase in cell proliferation (assessed by BrdU assay), cell density, and the size of cell nuclei. It is important to note that these changes should be always determined on the basis of a comparison with normal cells. Pathological changes at the tissue level include the formation of tumors without a clearly defined boundary, tumor invasion of neighboring tissues, an increase in angiogenesis, and disruption or destruction of normal tissues where cancer cells invade. The appearance of cancer cells at multiple locations suggests metastasis. Cancers are often

associated with rapid functional deterioration of involved organs. For instance, stomach cancer, when involving a large area, exhibits reduced capability of food digestion and rapid loss of body weight. Lung cancer is associated with difficulties in respiration. The ultimate consequence of cancer is the death of the involved organ. During the late stage, metastasis occurs in almost all cancer patients, often resulting in rapid failure of involved organ systems.

TREATMENT OF CANCERS

Conventional Treatment [25.1]

There are three major conventional approaches for the treatment of cancer: surgical removal, chemotherapy, and radiotherapy. *Surgical removal of cancers* is the most effective treatment when cancers are limited to a local area and are not spread to the surrounding tissues and lymph nodes. Early diagnosis of cancer is critical to the success of surgical treatment. Once cancers spread to neighboring tissues or metastasize to different organs, surgery is no longer effective. Chemotherapy and/or radiotherapy can be used for spread or metastasized cancers. *Chemotherapy* is an approach by which chemical agents are used to suppress the proliferation of cells, including cancer and normal cells. Since normal cells undergo a lower rate of division than cancer cells, chemical agents primarily affect cancer cells. A typical chemotherapeutic agent is 5-fluorouracil (5-FU), a uracil derivative that interrupts DNA synthesis and therefore suppresses cell division when incorporated into the genome of dividing cells. *Radiotherapy* is an approach used to treat cancers by exposing patients to radiation. While radiation destroys cancer cells and is effective for cancer treatment, it also induces normal cell injury and death.

Molecular Engineering Therapies

A number of molecular engineering strategies have been established and used for the treatment of cancers. These include the up-regulation of tumor suppressor genes, correction of mutant tumor suppressor genes, enhancement of anti-cancer immune responses, activation of tumor suppressor drugs, introduction of oncolytic viruses, and inhibition of growth-promoting genes by using antisense and siRNA oligonucleotides. A large number of investigations have been carried out to test these strategies in experimental models. Some of the strategies have been applied to preliminary clinical trials. Selected strategies are discussed as follows.

Overexpression of Tumor Suppressor Genes and Correction of Mutant Tumor Suppressor Genes [25.2]. As discussed on page 265, several proteins, including p16^{INK4}, p15^{INK4B}, p18^{INK4C}, p19^{INK4D}, p21, p27, and p57, are known to exert an inhibitory effect on cell division. These proteins may suppress the activity of cyclin D/CDK 4/CDK6, an important protein complex that regulates the progression of the cell division cycle from the G1 to S phase. Another protein, p53, suppresses cell division by activating p27. The overexpression of these proteins enhances the inhibitory effect on cell division and suppresses tumor cell growth. In particular, the p53 gene has been tested extensively. A large fraction of cancer patients exhibit mutant p53 gene, a potential factor contributing to the

initiation and development of cancers. Experimental investigations in animal models of cancers have shown that the transfer of the wildtype p53 gene into cancer tissues results in the suppression of cancer cell proliferation and reduction in cancer progression. Preliminary studies in human trials have demonstrated promising results for the therapeutic effect of the wildtype p53 gene. Other growth-inhibitory protein genes, as described above, can also serve as candidate genes for cancer therapy.

Enhancement of Anticancer Immune Responses [25.3]. Cancer cells express tumor antigens that can be recognized by the immune system under physiological conditions. The immune system is capable of destroying recognized cancer cells, as cancer cells are considered as foreign invaders. Such recognition and destruction activities are regulated by several signaling processes. When cancer cells form due to gene mutations, antigens expressed in the cancer cells can be recognized by antigen-presenting cells, which present the antigens to the T-helper cells. The T-helper cells produce and release cytokines, such as interleukin (IL)2, which activate cancer-specific T lymphocytes. The activated T lymphocytes can produce and release killer cytokines to suppress cancer growth and progression.

However, when the function of the immune system is suppressed, the immune system can no longer detect and destroy cancer cells, allowing cancer cells to proliferate and spread. Indeed, the suppression of the immune system may contribute to the initiation and development of cancers. Furthermore, cancer cells can produce and release cytokines that suppress the activity of host immune system and help cancer cells to escape from the immune surveillance and attack. Thus, a major approach for the treatment of cancers is to enhance the tumor-recognizing and suppressing functions of the host immune system. While it is difficult to achieve such a goal, several hypothetical strategies have been proposed and tested. These include the overexpression of T-lymphocyte activating factors, T-cell costimulating factors, and application of tumor-antigen vaccines.

The activity of the T lymphocytes can be stimulated by introducing several factors, such as human leukocyte antigen (HLA), major histocompatibility complex (MHC), cytokines (e.g., IL2 and interferon- γ) to target cells. The level of human leukocyte antigen is reduced in a large fraction of cancer patients. A molecule analogous to human leukocyte antigen is the major histocompatibility complex. The overexpression of the major histocompatibility complex gene in animal models of cancers induces activation of cytotoxic T cells and boosts anti-cancer responses. In human studies, the transfer of HLA-expressing vectors into melanoma cells induces the activation of cytotoxic T-cells and a reduction in tumor growth.

Several cytokines, including IL2, IL4, IL12 α (see Table 25.1 for specifics on IL12 α and CEA), granulocyte macrophage-colony stimulating factor (GM-CSF), and interferon- γ (see page 631 and the following table in this section for the characteristics of these factors), are known to promote the activation of T lymphocytes. The overexpression of these cytokines by gene transfer is a potential approach for the treatment of cancers. In particular, the IL2 gene has been tested in experimental models for anti-cancer effects. These studies have demonstrated that the overexpression of the IL2 gene activates cytotoxic T cells and reduces tumor growth in murine cancer models. Investigations in human trials have also shown promising results. Direct administration of cytokine proteins is another approach. However, cytokines undergo rapid degradation and the general administration of cytokines often induces toxic responses. Local gene transfer is an approach that can be used to overcome these problems.

TABLE 25.1. Characteristics of Carcinoembryonic Antigen and IL12 α *

Proteins	Alternative Names	Amino Acids	Molecular Weight (kDa)	Expression	Functions
Carcinoembryonic antigen	CEA, carcinoembryonic antigen-related antigen-related cell adhesion molecule 5 (CEACAM5), meconium antigen 100, CD66e antigen	702	77	Leukocyte, intestine, epithelial cells	Promoting cell survival and proliferation and enhancing cancer cell growth
IL12 α	IL12A, cytotoxic lymphocyte maturation factor, natural killer cell stimulatory factor chain (NKSF1), T-cell-stimulating factor	219	25	Dendritic cells	Inducing the expression of interferon (IFN) γ and promoting the differentiation of Th1 and Th2 cells

*Based on bibliography 25.3.

TABLE 25.2. Characteristics of B7.1*

Proteins	Alternative Names	Amino Acids	Molecular Weight (kDa)	Expression	Functions
B7.1	B7-1 antigen, B lymphocyte activation antigen B7-1, activation B7-1 antigen, CD28 antigen ligand 1 (CD28LG1), CD80 antigen	288	33	Liver, dendritic cells, mast cells, brain, blood vessel	Stimulating activation of T cells

*Based on bibliography 25.3.

Since the repression of the anticancer immune responses is considered a factor that contributes to tumorigenesis, the boosting of immune activities is a potential approach for the treatment of cancers. An effective approach for boosting immune activities is to deliver cancer-associated antigens to the host systems. Such antigens include viral analogues of tumor antigens, mutated oncogene proteins, and carcinoembryonic antigens. The genes of these proteins can be used to construct plasmids, known as recombinant vaccines, which can be used for gene transfer and for producing cancer-associated antigens. The host immune system is able to recognize these antigens and generate specific antibodies, which contribute to the anticancer activities. A typical tumor-associated antigen is the carcinoembryonic antigen, a glycoprotein receptor that is upregulated in certain types of cancer, such as colon cancer. The gene of the carcinoembryonic antigen can be inserted into a viral vector, such as the *Canary pox* virus, forming a recombinant DNA molecule. The overexpression of such a recombinant gene in a transgenic mouse model results in the production of antibodies against the carcinoembryonic antigen, in association with the enhancement of T-cell activities against tumor cells that express the carcinoembryonic antigen. Clinical investigations in patients with colorectal cancer have shown promising results, including the generation of antibodies against the carcinoembryonic antigen and activation of the cytotoxic T cells.

The activation of cytotoxic T cells can be induced by exposure to cancer cell antigens. The activity of the cytotoxic T cells can be boosted by costimulating factors such as the B7.1 protein (see Table 25.2). In certain types of cancer, the expression of costimulating factors is repressed, a potential factor that reduces the anticancer immune responses. Thus, a strategy for enhancing the activity of the cytotoxic T cells is to deliver cancer cell antigens and costimulating factors together. A recombinant gene can be constructed with the carcinoembryonic antigen gene and the costimulating factor B7.1 gene. Such a gene construct has been applied to human patients with adenocarcinoma in a preliminary clinical trial. This study has demonstrated that the overexpression of the recombinant gene is associated with activation of T cells specific to the carcinoembryonic antigen. The application of the costimulating factor gene enhances the anticancer immune reactions.

Activation of Tumor-Suppressing Prodrugs [25.4]. Chemotherapy is an effective approach for the inhibition of cancer metastasis. However, most chemotherapeutic agents destroy not only cancer cells, but also normal cells, significantly repressing the activity of the immune system. It is desired to introduce chemotherapeutic agents that are effective only in targeted cancer cells without influencing the normal cells. Synthetic deoxynucleosides may serve as such chemotherapeutic agents. A typical agent is ganciclovir, an analogue of deoxyguanosine. Ganciclovir can be phosphorylated by the herpes simplex virus tyrosine kinase to form ganciclovir triphosphate, a deoxynucleotide that can be incorporated into DNA during DNA synthesis in place of dGTP. The incorporation of the ganciclovir deoxynucleotide induces the termination of DNA synthesis, thus arresting cell division in the S phase. Ganciclovir without phosphorylation is not effective. Based on such a feature, ganciclovir can be administered through blood injection, while herpes simplex virus tyrosine kinase gene can be delivered to local target cancer cells. With the expression of the viral tyrosine kinase gene in the local cancer cells, ganciclovir can only be phosphorylated in the cancer cells, thus repressing DNA synthesis in cancers but not in normal cells. Experimental investigations have demonstrated that in animal models of

colon cancer, general ganciclovir administration with local delivery of the herpes simplex viral tyrosine kinase gene results in a significant reduction in tumor growth. Clinical investigations with a similar approach have also shown promising results for the treatment of human colorectal adenocarcinoma.

Another tumor-suppressing prodrug is 5-fluorocytosine, a cytosine derivative. This agent is relatively inactive in its natural form. A cytosine-specific enzyme called cytosine deaminase, which is found in bacteria and fungi, can convert 5-fluorocytosine to 5-fluorouracil, a potent anti-cancer agent that blocks the methylation reaction of deoxyuridylic acid to thymidylic acid and thus interferes with DNA synthesis. A blood injection of 5-fluorocytosine, together with a delivery of a bacterial cytosine deaminase gene into local target cancers can induce local conversion of 5-fluorocytosine to 5-fluorouracil, resulting in the repression of cancer growth with a minimal influence on normal cell function. Such an approach has been shown to effectively inhibit cancer proliferation in animal models as well as in human patients with colorectal cancer.

Application of Oncolytic Viruses [25.5]. Several types of viruses, including adenovirus and herpes simplex virus, can be genetically modified to establish the capability of lysing cancer cells, but not normal cells. A mutant adenovirus has been created by removing the E1B segment of the viral genome, resulting in the deficiency of the E1B-55kDa viral protein. When delivered to cancer cells, the mutant adenoviruses can replicate in cancer cells lacking p53 and then lyse these cells. Clinical investigations have shown that the delivery of mutant adenoviruses into metastatic tumors results in cancer cell death. However, controversial results have been reported regarding the target specificity of the mutant adenoviruses. Further investigations are necessary to confirm the preliminary discoveries.

Herpes simplex virus is another type of virus that can be modified to establish cancer cell-specific lysing activity. The removal of selective genes, including the tyrosine kinase gene or the ribonucleotide reductase, from the herpes simplex virus results in mutant viruses that can replicate in dividing cells. Since cancer cells undergo a much higher rate of division compared to normal cells, the transfer of the mutant herpes simplex viruses into cancer cells results in the lysis of these cells. Experimental studies have demonstrated selective oncolytic activity of herpes simplex virus in animal colon cancer models.

Application of Antisense Oligonucleotides and siRNA [25.6]. The initiation and development of cancers involve the upregulation of oncogenic genes, such as *abl* (protein tyrosine kinase), *raf* (protein serine/threonine kinase), *ras* (GTP-binding protein), *sis* (platelet-derived growth factor B chain), and *src* (Src protein tyrosine kinase). The expression of other genes that encode mitogenic proteins also contributes to carcinogenesis. The suppression of carcinogenic gene expression is an effective approach for the treatment of cancers. A typical approach for such a purpose is the administration of antisense oligonucleotides specific to a target mRNA molecule. Antisense oligonucleotides are short fragments of nucleotides with a length of 20–40 base pairs and can hybridize to complementary mRNA in the cytoplasm, blocking the translation of proteins. These fragments can be directly delivered to target cells for therapeutic purposes. An alternative method is to transfer a gene construct that encodes a fragment of antisense deoxynucleotides,

which can be expressed and exerts antisense effects. Furthermore, the transfection of target cancer cells with siRNAs specific to proliferative mRNAs can effectively suppress the proliferation of cancer cells (Fig. 25.1).

Application of Combined Therapeutic Approaches [25.7]. The treatment of cancers is a challenging task. Often, the application of a single approach as described above may not be effective, especially in the presence of cancer metastasis. The combination of anti-cancer therapeutic methods may provide an alternative means for the treatment of cancers. Various combinations have been tested in experimental and clinical investigations. Exam-

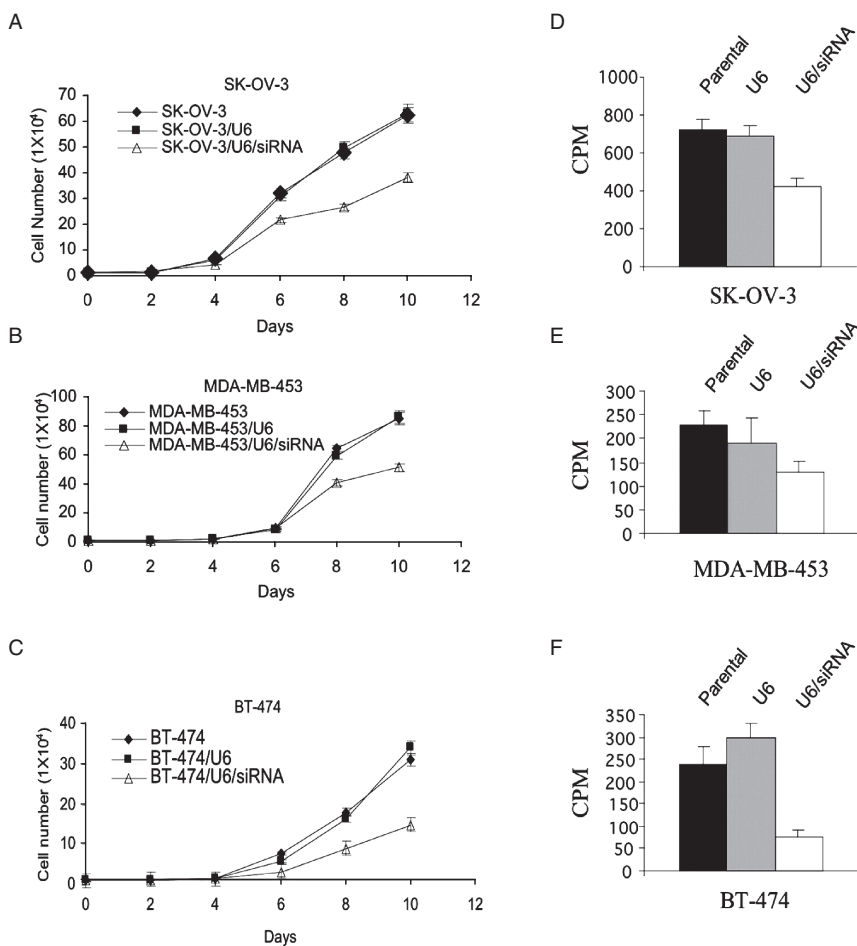


Figure 25.1. Cell proliferation, measured by cell counts (A–C) and [3 H]thymidine uptake (D–F) of SK-OV-3, MDA-MB-453, and BT-474 cells with and without transfection with Her-2/neu siRNA. Note that the *Her-2/neu* gene is an oncogene that is overexpressed in ~30% of breast and ovarian cancer cases. The expression of this gene often indicates a poor prognosis. CPM: counts per minute. (Reprinted with permission from Yang G et al: Inhibition of breast and ovarian tumor growth through multiple signaling pathways by using retrovirus-mediated small interfering RNA against Her-2/neu gene expression, *J Biol Chem* 279:4339–45, copyright 2004.)

ples include the combination of oncolytic viruses and anticancer prodrugs, the combination of radiotherapy with oncolytic viruses, and the combination of tumor-suppressing genes and immune response activators. These combination approaches have been shown to be more effective compared to single approaches.

BIBLIOGRAPHY

25.1. Classification, Pathogenesis, and Treatment of Cancer

- Brinkman BM, Wong DT: Disease mechanism and biomarkers of oral squamous cell carcinoma, *Curr Opin Oncol* 18(3):228–33, May 2006.
- Kleinsmith LJ: *Principles of Cancer Biology*, Pearson Benjamin Cummings, San Francisco, 2006.
- Schiff D, O'Neill B: *Principles of Neurooncology*, McGraw-Hill, New York, 2005.
- Vokes EE, Golomb HM: *Oncologic Therapies*, Springer, Berlin, 2003.
- Pitot HC: *Fundamentals of Oncology*, 4th ed, Marcel Dekker, New York, 2002.
- Stone RM: *Harrison's Principles of Internal Med*, 15th ed, McGraw-Hill, New York, 2001.
- Schneider AS, Szanto PA: *Pathology*, 3rd ed, Lippincott Williams & Wilkins, Philadelphia, 2006.
- McCance KL, Huether SE: *Pathophysiology: The Biologic Basis for Disease in Adults & Children*, 5th ed, Elsevier Mosby, St. Louis, 2006.
- Porth CM: *Pathophysiology: Concepts of Altered Health States*, 7th ed, Lippincott Williams & Wilkins, Philadelphia, 2005.
- Frazier MS, Drzymkowski JW: *Essentials of Human Diseases and Conditions*, 3rd ed, Elsevier Saunders, St Louis, 2004.

25.2. Overexpression of Tumor Suppressor Genes and Correction of Mutant Tumor Suppressor Genes

- Harris MP, Sutjipto S, Wills KN et al: Adenovirus-mediated p53 gene transfer inhibits growth of human tumor cells expressing mutant p53 protein, *Cancer Gene Ther* 3:121–30, 1996.
- Bouvet M, Ellis LM, Nishizaki M et al: Adenovirus-mediated wild-type p53 gene transfer down-regulates vascular endothelial growth factor expression and inhibits angiogenesis in human colon cancer, *Cancer Res* 58:2288–92, 1998.
- Cohen AM, Kemeny NE, Kohne CH, Wils J, de Takats PG et al: Is intra-arterial chemotherapy worthwhile in the treatment of patients with unresectable hepatic colorectal cancer metastases? *Eur J Cancer* 32A:2195–205, 1996.
- Nguyen DM, Spitz FR, Yen N, Cristiano RJ, Roth JA: Gene therapy for lung cancer: Enhancement of tumor suppression by a combination of sequential systemic cisplatin and adenovirus-mediated p53 gene transfer, *J Thor Cardiovasc Surg* 112:1372–6, 1376–7, 1996.

25.3. Enhancement of Anti-Cancer Immune Responses

- Todryk SM, Chong H, Vile RG, Pandha H, Lemoine NR: Can immunotherapy by gene transfer tip the balance against colorectal cancer? *Gut* 43:445–9, 1998.
- Bodey B, Bodey Jr B, Siegel SE, Kaiser HE: Failure of cancer vaccines: The significant limitations of this approach to immunotherapy, *Anticancer Res* 20:2665–76, 2000.
- Ostrand-Rosenberg S, Thakur A, Clements V: Rejection of mouse sarcoma cells after transfection of MHC class II genes, *J Immunol* 144:4068–71, 1990.

- Nabel GJ, Nabel EG, Yang ZY et al: Direct gene transfer with DNA-liposome complexes in melanoma: Expression, biologic activity, and lack of toxicity in humans, *Proc Natl Acad Sci USA* 90:11307–11, 1993.
- Gonzalez R, Atkins M, Schwarzenberger P et al: Phase II trial of HLA-B7 plasmid DNA/lipid (Allovectin®) immunotherapy in patients with metastatic melanoma, *Proc Am Soc Clin Oncol* 20:1007, 2001.
- Rubin J, Galanis E, Pitot HC et al: Phase I study of immunotherapy of hepatic metastases of colorectal carcinoma by direct gene transfer of an allogeneic histocompatibility antigen, HLA-B7, *Gene Ther* 4:419–25, 1997.

Carcinoembryonic Antigen (CEA)

- Brandriff BF, Gordon LA, Tynan KT, Olsen AS, Mohrenweiser HW et al: Order and genomic distances among members of the carcinoembryonic antigen (CEA) gene family determined by fluorescence in situ hybridization, *Genomics* 12:773–9, 1992.
- Nishi M, Inazawa J, Inoue K, Nakagawa H, Taniwaki M et al: Regional chromosomal assignment of carcinoembryonic antigen gene (CEA) to chromosome 19 at band q13.2, *Cancer Genet Cytogenet* 54:77–81, 1991.
- Thompson JA, Pande H, Paxton RJ, Shively L, Padma A et al: Molecular cloning of a gene belonging to the carcinoembryonic antigen gene family and discussion of a domain model, *Proc Natl Acad Sci USA* 84:2965–9, 1987.
- Willcocks TC, Craig SP, Craig IW: Assignment of the coding sequence for carcinoembryonic antigen (CEA) and normal cross-reacting antigen (NCA) to human chromosome 19q13, *Ann Hum Genet* 53:141–8, 1989.
- Zimmer R, Thomas P: Mutations in the carcinoembryonic antigen gene in colorectal cancer patients: Implications on liver metastasis, *Cancer Res* 61:2822–6, 2001.
- Zimmermann W, Ortlieb B, Friedrich R, von Kleist S: Isolation and characterization of cDNA clones encoding the human carcinoembryonic antigen reveal a highly conserved repeating structure, *Proc Natl Acad Sci USA* 84:2960–4, 1987.

IL12

- Kass E, Schlom J, Thompson J, Guadagni F, Graziano P et al: Induction of protective host immunity to carcinoembryonic antigen (CEA), a self-antigen in CEA transgenic mice, by immunizing with a recombinant vaccinia-CEA virus, *Cancer Res* 59:676–83, 1999.
- Tsang KY, Zaremba S, Nieroda CA, Zhu MZ, Hamilton JM et al: Generation of human cytotoxic T cells specific for human carcinoembryonic antigen epitopes from patients immunized with recombinant vaccinia-CEA vaccine, *J Natl Cancer Inst* 87:982–90, 1995.
- Conry RM, Khazaeli MB, Saleh MN et al: Phase I trial of a recombinant vaccinia virus encoding carcinoembryonic antigen in metastatic adenocarcinoma: Comparison of intradermal versus subcutaneous administration, *Clin Cancer Res* 5:2330–7, 1999.
- Conry RM, Allen KO, Lee S, Moore SE, Shaw DR et al: Human autoantibodies to carcinoembryonic antigen (CEA) induced by a vaccinia-CEA vaccine, *Clin Cancer Res* 6:34–41, 2000.
- Hodge JW, McLaughlin JP, Kantor JA, Schlom J: Diversified prime and boost protocols using recombinant vaccinia virus and recombinant non-replicating avian pox virus to enhance T-cell immunity and antitumor responses, *Vaccine* 15:759–68, 1997.
- Marshall JL, Hawkins MJ, Tsang KY et al: Phase I study in cancer patients of a replication-defective avipox recombinant vaccine that expresses human carcinoembryonic antigen, *J Clin Oncol* 17:332–7, 1999.
- Zhu MZ, Marshall J, Cole D, Schlom J, Tsang KY: Specific cytolytic T-cell responses to human CEA from patients immunized with recombinant avipox-CEA vaccine, *Clin Cancer Res* 6: 24–33, 2000.

B7.1

- Freeman GJ, Disteche CM, Gribben JG, Adler DA, Freedman AS et al: The gene for B7, a costimulatory signal for T-cell activation, maps to chromosomal region 3q13.3-3q21, *Blood* 79:489–94, 1992.
- Reiser J, von Gersdorff G, Loos M, Oh J, Asanuma K et al: Induction of B7-1 in podocytes is associated with nephrotic syndrome, *J Clin Invest* 113:1390–7, 2004.
- Selvakumar A, Mohanraj BK, Eddy RL, Shows TB, White PC et al: Genomic organization and chromosomal location of the human gene encoding the B-lymphocyte activation antigen B7, *Immunogenetics* 36:175–81, 1992.
- Shah R, Banks K, Patel A, Dogra S, Terrell R et al: Intense expression of the B7-2 antigen presentation coactivator is an unfavorable prognostic indicator for differentiated thyroid carcinoma of children and adolescents, *J Clin Endocr Metab* 87:4391–7, 2002.
- Stamper CC, Zhang Y, Tobin JF, Erbe DV, Ikemizu S et al: Crystal structure of the B7-1/CTLA-4 complex that inhibits human immune responses, *Nature* 410:608–11, 2001.
- Horig H, Lee DS, Conkright W et al: Phase I clinical trial of a recombinant canarypoxvirus (ALVAC) vaccine expressing human carcinoembryonic antigen and the B7.1 co-stimulatory molecule, *Cancer Immunol Immunother* 49:504–14, 2000.
- Fakhrai H, Shawler DL, Gjerset R et al: Cytokine gene therapy with interleukin-2-transduced fibroblasts: Effects of IL-2 dose on anti-tumor immunity, *Hum Gene Ther* 6:591–601, 1995.
- Sobol RE, Shawler DL, Carson C et al: Interleukin 2 gene therapy of colorectal carcinoma with autologous irradiated tumor cells and genetically engineered fibroblasts: A phase I study, *Clin Cancer Res* 5:2359–65, 1999.
- Suminami Y, Elder EM, Lotze MT, Whiteside TL: In situ interleukin-4 gene expression in cancer patients treated with genetically modified tumor vaccine, *J Immunother Emphasis Tumor Immunol* 17:238–48, 1995.
- Schmidt-Wolf IG, Finke S, Trojanek B et al: Phase I clinical study applying autologous immunological effector cells transfected with the interleukin-2 gene in patients with metastatic renal cancer, colorectal cancer and lymphoma, *Br J Cancer* 81:1009–16, 1999.
- Rochlitz CF, Jantschke P, Bongartz G et al: Gene therapy with cytokine-transfected xenogeneic cells in metastatic tumors, *Adv Exp Med Biol* 451:531–7, 1998.
- Galanis E, Hersh EM, Stopeck AT et al: Immunotherapy of advanced malignancy by direct gene transfer of an interleukin-2 DNA/DMRIE/DOPE lipid complex: Phase I/II experience, *J Clin Oncol* 17:3313–23, 1999.
- Geutskens SB, van der Eb MM, Plomp AC et al: Recombinant adenoviral vectors have adjuvant activity and stimulate T cell responses against tumor cells, *Gene Ther* 7:1410–6, 2000.
- Human protein reference data base, Johns Hopkins University and the Institute of Bioinformatics, at <http://www.hprd.org/protein>.

25.4. Activation of Tumor-Suppressing Pro-Drugs

- Link Jr CJ, Levy JP, McCann LZ, Moorman DW: Gene therapy for colon cancer with the herpes simplex thymidine kinase gene, *J Surg Oncol* 64:289–94, 1997.
- Sung MW, Yeh HC, Thung SN et al: Intratumoral adenovirus-mediated suicide gene transfer for hepatic metastases from colorectal adenocarcinoma: Results of a phase I clinical trial, *Mol Ther* 4:182–91, 2001.
- Boucher PD, Im MM, Freytag SO, Shewach DS: A novel mechanism of synergistic cytotoxicity with 5-fluorocytosine and ganciclovir in double suicide gene therapy, *Cancer Res* 66(6):3230–7, 2006.
- Kuriyama S, Kikukawa M, Masui K et al: Cytosine deaminase/5-fluorocytosine gene therapy can induce efficient anti-tumor effects and protective immunity in immunocompetent mice but not in athymic nude, *Int J Cancer* 81:592–7, 1999.

- Huber BE, Austin EA, Richards CA, Davis ST, Good SS: Metabolism of 5-fluorocytosine to 5-fluorouracil in human colorectal tumor cells transduced with the cytosine deaminase gene: Significant antitumor effects when only a small percentage of tumor cells express cytosine deaminase, *Proc Natl Acad Sci USA* 91:8302–6, 1994.
- Crystal RG, Hirschowitz E, Lieberman M et al: Phase I study of direct administration of a replication deficient adenovirus vector containing the E. coli cytosine deaminase gene to metastatic colon carcinoma of the liver in association with the oral administration of the pro-drug 5-fluorocytosine, *Hum Gene Ther* 8:985–1001, 1997.

25.5. Application of Oncolytic Viruses

- Chernajovsky Y, Layward L, Lemoine N: Fighting cancer with oncolytic viruses, *Br Med J* 332(7534):170–2, 2006.
- Aghi M, Martuza RL: Oncolytic viral therapies—the clinical experience, *Oncogene* 24(52):7802–16, 2005.
- Working PK, Lin A, Borellini F: Meeting product development challenges in manufacturing clinical grade oncolytic adenoviruses, *Oncogene* 24(52):7792–801, 2005.
- Mathis JM, Stoff-Khalili MA, Curiel DT: Oncolytic adenoviruses—selective retargeting to tumor cells, *Oncogene* 24(52):7775–91, 2005.
- Ko D, Hawkins L, Yu DC: Development of transcriptionally regulated oncolytic adenoviruses, *Oncogene* 24(52):7763–74, 2005.
- Shmulevitz M, Marcato P, Lee PW: Unshackling the links between reovirus oncolysis, Ras signaling, translational control and cancer, *Oncogene* 24(52):7720–8, 2005.
- Barber GN: VSV-tumor selective replication and protein translation, *Oncogene* 24(52):7710–9, 2005.
- Mohr I: To replicate or not to replicate: Achieving selective oncolytic virus replication in cancer cells through translational control, *Oncogene* 24(52):7697–709, 2005.
- O’Shea CC: Viruses—seeking and destroying the tumor program, *Oncogene* 24(52):7640–55, 2005.
- Chou J, Roizman B: The gamma 1(34.5) gene of herpes simplex virus 1 precludes neuroblastoma cells from triggering total shutoff of protein synthesis characteristic of programmed cell death in neuronal cells, *Proc Natl Acad Sci USA* 89:3266–70, 1992.
- Mineta T, Rabkin SD, Yazaki T, Hunter WD, Martuza RL: Attenuated multi-mutated herpes simplex virus-1 for the treatment of malignant gliomas, *Nat Med* 1:938–43, 1995.
- Walker JR, McGeagh KG, Sundaresan P, Jorgensen TJ, Rabkin SD et al: Local and systemic therapy of human prostate adenocarcinoma with the conditionally replicating herpes simplex virus vector G207, *Hum Gene Ther* 10:2237–43, 1999.
- Delman KA, Bennett JJ, Zager JS et al: Effects of preexisting immunity on the response to herpes simplex-based oncolytic viral therapy, *Hum Gene Ther* 11:2465–72, 2000.
- Wildner O, Blaese RM, Morris JC: Therapy of colon cancer with oncolytic adenovirus is enhanced by the addition of herpes simplex virus-thymidine kinase, *Cancer Res* 59:410–3, 1999.
- Bischoff JR, Kirn DH, Williams A et al: An adenovirus mutant that replicates selectively in p53-deficient human tumor cells, *Science* 274:373–6, 1996.
- Rothmann T, Hengsternmann A, Whitaker NJ, Scheffner M, zur Hausen H: Replication of ONYX-015, a potential anticancer adenovirus, is independent of p53 status in tumor cells, *J Virol* 72:9470–8, 1998.
- Ries SJ, Brandts CH, Chung AS et al: Loss of p14ARF in tumor cells facilitates replication of the adenovirus mutant dl1520 (ONYX-015), *Nat Med* 6:1128–33, 2000.

- Reid T, Galanis E, Abbruzzese J et al: Intra-arterial administration of a replication-selective adenovirus (dl1520) in patients with colorectal carcinoma metastatic to the liver: A phase I trial, *Gene Ther* 8:1618–26, 2001.
- Habib NA, Sarraf CE, Mitry RR et al: E1B-deleted adenovirus (dl1520) gene therapy for patients with primary and secondary liver tumors, *Hum Gene Ther* 12:219–26, 2001.
- Khuri FR, Nemunaitis J, Ganly I et al: A controlled trial of intratumoral ONYX-015, a selectively-replicating adenovirus, in combination with cisplatin and 5-fluorouracil in patients with recurrent head and neck cancer, *Nat Med* 6:879–85, 2000.
- Heise C, Hermiston T, Johnson L et al: An adenovirus E1A mutant that demonstrates potent and selective systemic anti-tumoral efficacy, *Nat Med* 6:1134–9, 2000.

25.6. Application of Antisense Oligodeoxynucleotides

- Tortora G, Caputo R, Damiano V et al: A novel MDM2 anti-sense oligonucleotide has anti-tumor activity and potentiates cytotoxic drugs acting by different mechanisms in human colon cancer, *Int J Cancer* 88:804–9, 2000.
- Berg RW, Werner M, Ferguson PJ et al: Tumor growth inhibition in vivo and G2/M cell cycle arrest induced by antisense oligodeoxynucleotide targeting thymidylate synthase, *J Pharmacol Exp Ther* 298:477–84, 2001.
- Nishizuka I, Ichikawa Y, Ishikawa T et al: Matrilysin stimulates DNA synthesis of cultured vascular endothelial cells and induces angiogenesis in vivo, *Cancer Lett* 173:175–82, 2001.
- Tamm I, Dorken B, Hartmann G: Antisense therapy in oncology: New hope for an old idea? *Lancet* 358:489–97, 2001.

25.7. Application of Combined Therapeutic Approaches

- Carroll NM, Chase M, Chiocca EA, Tanabe KK: The effect of ganciclovir on herpes simplex virus-mediated oncolysis, *J Surg Res* 69:413–7, 1997.
- Rogulski KR, Wing MS, Paielli DL, Gilbert JD, Kim JH et al: Double suicide gene therapy augments the antitumor activity of a replication-competent lytic adenovirus through enhanced cytotoxicity and radiosensitization, *Hum Gene Ther* 11:67–76, 2000.
- Andreansky S, He B, van Cott J et al: Treatment of intracranial gliomas in immunocompetent mice using herpes simplex viruses that express murine interleukins, *Gene Ther* 5:121–30, 1998.
- Advani SJ, Sibley GS, Song PY et al: Enhancement of replication of genetically engineered herpes simplex viruses by ionizing radiation: A new paradigm for destruction of therapeutically intrac-table tumors, *Gene Ther* 5:160–5, 1998.
- Shimada H, Shimizu T, Ochiai T, Liu TL, Sashiyama H et al: Preclinical study of adenoviral p53 gene therapy for esophageal cancer, *Surg Today* 31(7):597–604, 2001.