A Detection Method for Thermoresistant Bacteria

Abstract: In cultivating bacteria, time is regarded as a signal factor. Since a certain number of bacteria exist before starting, we used a reference-point proportional equation with the ninth hour from the start as a reference time. On the other hand, because the real number of bacteria was unknown, the numbers diluted to 1/5 and 1/25 were chosen as signal factors to be used for measurement. In other words, using dilution ratio as the signal, we calculated an SN ratio whose true values were unknown.

1. Introduction

Thermoresistant bacteria form heat-resisting spores in the body that survive in the food after cooking and rot heat-treated food. In order to detect a specific bacterium causing food poisoning, we developed a method of detecting poison produced by a bacterium or technique using an antigen-antibody reaction. However, since a heat-resistant bacterium is a common decomposing bacterium, a method of detecting cultivated bacteria is widely used. In this case, a key issue was to accelerate bacteria as quickly as possible and to detect them accurately. If the speed of occurrence of bacteria is slow, accuracy in detection is reduced. In contrast, poor accuracy of detection leads to a larger error in confirmation of occurrence. This relationship between a measuring method and a measured result cannot be separated in technological development, and each should be analyzed individually. Whereas cultivation of bacteria is considered a growth-related phenomenon, detection is a measurement- or SN ratio-related issue. In our study we dealt with both in a single experiment.

In cultivating bacteria, time is regarded as a signal factor (Figure 1). Since a certain number of bacteria exist before starting, we used a reference-point proportional equation with the ninth hour from the start as a reference time. On the other hand, because the real number of bacteria was unknown, the numbers diluted to 1/5 and 1/25 were chosen as signal factors to be used for measurement. In other words, using dilution ratio as the signal, we calculate an SN ratio whose true values were unknown (Figure 2). In addition, on the assumption that the pH of the food had already been adjusted to control microbes, it was considered as the noise factor:

 N_1 : diluted solution, pH 7

 N_2 : diluted solution, pH 3

Following our objective of study, we proceeded with the following data analysis:

- 1. To perform parameter design for bacteria cultivation, we calculated the SN ratio of the reference-point proportional equation using time as a signal factor (analysis 1).
- 2. By computing the SN ratio for bacteria detection at each point of time, we used bacteria dilution ratio as a signal factor (analysis 2).

Using the results obtained from both, we chose the optimal configuration for cultivation and detection methods for bacteria.

SN Ratio for Cultivation Conditions for Bacteria (Analysis 1)

As a data example, the data in the first row of an L_{18} orthogonal array are shown in Table 1. To

Case 8

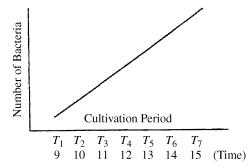


Figure 1 Cultivation of bacteria using time as the signal

compute the SN ratio using a reference-point proportional equation whose reference point is set to T_1 , we created an auxiliary table (Table 2).

Total variation:

$$S_T = 79.5^2 + 147.5^2 + \dots + 7.5^2 + 8.5^2$$

= 603,265 (1)

Effective divider:

$$r = 0^2 + 1^2 + \dots + 6^2 = 91 \tag{2}$$

Linear equation:

$$L_1 = (0)(79.5) + \dots + (6)(346.5) = 6178.5$$

$$L_2 = 2965.5$$
 $L_3 = 2018.0$

$$L_4 = 167.0$$
 $L_5 = 590.5$ $L_6 = 133.5$ (3)

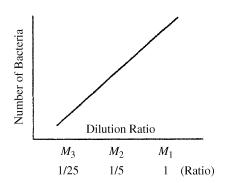


Figure 2

Measurement of bacteria using dilution of bacteria as the signal

Table	1
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Measured data of number of bacteria in experiment 1

		M ₁	<i>M</i> ₂	M ₃
<i>T</i> ₁	$egin{array}{c} N_1 \ N_2 \end{array}$	496 337	119 35	11 8
<i>T</i> ₂	$egin{array}{c} N_1 \ N_2 \end{array}$	564 425	137 50	23 12
<i>T</i> ₃	$egin{array}{c} N_1 \ N_2 \end{array}$	612 480	153 63	27 13
<i>T</i> ₄	$egin{array}{c} N_1 \ N_2 \end{array}$	663 525	164 76	34 14
T ₅	$egin{array}{c} N_1 \ N_2 \end{array}$	710 558	171 86	40 15
<i>T</i> ₆	$egin{array}{c} N_1 \ N_2 \end{array}$	746 578	180 92	41 17
<i>T</i> ₇	$egin{array}{c} N_1 \ N_2 \end{array}$	763 605	186 96	41 18

Table 2

Calibrated data for reference-point proportional equation

		M ₁	<i>M</i> ₂	M ₃
0	$egin{array}{c} N_1 \ N_2 \end{array}$	79.5 -79.5	42.0 -42.0	$\begin{array}{c} 1.5 \\ -1.5 \end{array}$
1	$egin{array}{c} N_1 \ N_2 \end{array}$	147.5 8.5	60.0 -27.0	13.5 2.5
2	$egin{array}{c} N_1 \ N_2 \end{array}$	195.5 63.5	76.0 -14.0	17.5 3.5
3	$egin{array}{c} N_1 \ N_2 \end{array}$	246.5 108.5	87.0 -1.0	24.5 4.5
4	$egin{array}{c} N_1 \ N_2 \end{array}$	293.5 141.5	94.0 9.0	30.5 5.5
5	$egin{array}{c} N_1 \ N_2 \end{array}$	329.5 161.5	103.0 15.0	31.5 7.5
6	$egin{array}{c} N_1 \ N_2 \end{array}$	346.5 188.5	109.0 19.0	31.5 8.5
Linear equation	$egin{array}{c} N_1 \ N_2 \end{array}$	L_1 L_2	L ₃ L ₄	L ₅ L ₆

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Variation of proportional term:

$$S_{\beta} = \frac{(L_1 + L_2 + L_3 + L_4 + L_5 + L_6)^2}{6r}$$

= 266,071.08 (4)

Variation of difference between proportional terms by dilution:

$$S_{M\beta} = \frac{(L_1 + L_2)^2 + (L_3 + L_4)^2 + (L_5 + L_6)^2}{2r} - S_{\beta}$$

= 222,451.65 (5)

Variation of differences between proportional terms due to noise:

$$S_{N\beta} = \frac{(L_1 + L_3 + L_5)^2 + (L_2 + L_4 + L_6)^2}{3r} - S_{\beta}$$

= 55,826.82 (6)

Error variation:

$$S_e = S_T - S_\beta - S_{M\beta} - S_{N\beta} = 58,915.45$$
 (7)

Error variance:

$$V_e = \frac{S_e}{38} = 1550.41 \tag{8}$$

Total variance:

$$V_N = \frac{S_{N\beta} + S_e}{39} \times 2942.11$$
(9)

SN ratio:

$$\eta = 10 \log \frac{(S_{\beta} - V_{\ell})/2r}{V_N} = -3.06 \qquad (10)$$

Sensitivity:

$$S = 10 \log \frac{S_{\beta} - V_{e}}{2r} = 31.62 \tag{11}$$

3. SN Ratio for Measurement of Number of Bacteria (Analysis 2)

Using the data of time T_1 in Table 1, we calculated the SN ratio for using the bacteria dilution ratio. Table 3 summarizes each datum divided by 416.5, the average number of bacteria at M_1 .

Table 3

Calibrated data

		1	0.2	0.04
<i>T</i> ₁	$egin{array}{c} N_1 \ N_2 \end{array}$	1.191 0.809	0.286 0.084	0.026 0.019

Total variation:

$$S_T = 1.191^2 + 0.286^2 + \dots + 0.019^2 = 2.163$$
 (12)

Effective divider:

$$r = 1^2 + 0.2^2 + 0.04^2 = 1.042 \tag{13}$$

Linear equations:

$$L_1 = (1)(1.191) + \dots + (0.04)(0.026) = 1.249$$

$$L_2 = 0.827$$
(14)

Variation of proportional term:

$$S_{\beta} = \frac{(L_1 + L_2)^2}{2r} = 2.068 \tag{15}$$

Variation of difference between proportional terms:

$$S_{N\beta} = \frac{L_1^2 + L_2^2}{r} - S_{\beta} = 0.086 \tag{16}$$

Error variation:

$$S_e = S_T - S_\beta - S_{N\beta} = 0.009 \tag{17}$$

Error variance:

$$V_e = \frac{S_e}{4} = 0.002 \tag{18}$$

Total variance:

$$V_N = \frac{S_{N\beta} + S_e}{5} = 0.019 \tag{19}$$

SN ratio:

$$\eta = 10 \log \frac{(S_{\beta} - V_{\ell})/2r}{V_N} = 17.216 \qquad (20)$$

Sensitivity:

$$S = 10 \log \frac{S_{\beta} - V_{e}}{2r} = -0.04 \tag{21}$$

We also computed each value for T_2 to T_7 and then proceeded with the same calculation for experiments 2 to 18.

Optimization of Cultivation and Detection Methods and Confirmatory Experiment

Table 4 shows the control factors for the design of experiments on cultivation and detection of bacteria. Type of bacterium, A, was allocated as an indicative factor. For control factors, we chose type of dilution solution, B; type of medium, D; and amounts of elements E to H, which are assumed to facilitate growth of bacteria (added to medium). On the other hand, bench time C, measured in terms of the number of days, with bacteria maintained at 10°C after being heated up.

As below, we showed the response graphs for measurement of the number of bacteria (analysis 2). Figure 3 shows the response graphs for factor A only. Since analysis is conducted for each time level, there will be response graphs of T_1 to T_7 for other factors.

We obtained the following identical optimal configurations for analyses 1 and 2:

Under both optimal and current configurations, we performed a confirmatory experiment. The results are shown in Table 5.

5. Results of Experiment

According to the results of the confirmatory experiment shown in Table 5, we discovered that satisfactory reproducibility in the gain of the SN ratio cannot be obtained. However, since the trend toward increasing SN ratio with respect to time in the results of analysis 2 was similar to that of the estimation, we concluded that our experimental results were fairly reliable. On the other hand, a number of peaks and V-shapes in the response graphs for the control factors were regarded as problems to be solved in a future study.

Despite several remaining problems, the following improvements were achieved:

1. We reduced the experimentation time by two days because the bench time allocated as one of the control factors was not necessary. More-

Table 4

Control and indicative factors and levels

		Level			
	Factor	1	2	3	
<i>A</i> :	type of bacterium (<i>B. subtilis</i>)	A	В	—	
В:	dilution solution	Sterile water	Phosphate buffer solution	Peptone– phosphate buffer solution	
<i>C</i> :	bench time (days)	0	1	2	
D:	type of medium	Standard	Glucose tryptone	Trypticase soy	
<i>E</i> :	amount of catalase (μg/mL)	0	50	100	
F:	amount of lysozyme (µg/mL)	0	0.01	0.1	
G:	amount of sodium pyruvate (%)	0.00	0.10	0.20	
H:	amount of alanine (μ g/mL)	0	20	40	

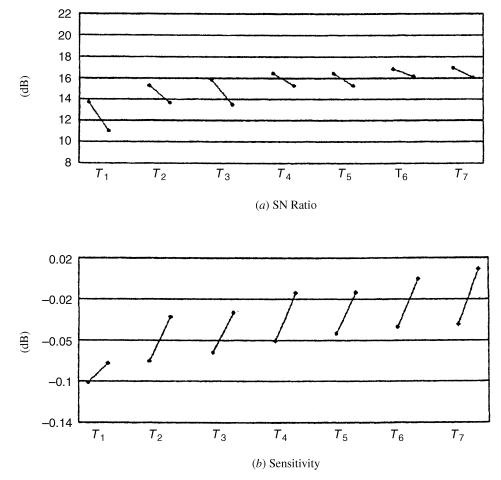


Figure 3 Analysis 2: Response graphs for factor *A*

over, whereas five days are used as a cultivation period in our current process, we found such a long-time cultivation not meaningful for experimentation. Therefore, we finished an inspection that normally takes one week in only two days.

2. The improvement above contributed not only to reducing the cost but also to taking countermeasures more swiftly. If we can detect the occurrence of bacteria sooner, we can more retrieve defective products immediately, and thereby alleviate the risk of inflicting damage on customers.

3. For detectability of bacteria, we obtained improvement of 15 dB in the SN ratio (1/32 of the current variability) regarding a cultivation condition of bacteria when time was selected as a signal factor, whereas the bacteria dilution ratio was approximately 21 dB (1/122 of the current variability) after 15 hours.

The resulting economic benefits were computed as follows:

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Table 5

Confirmatory experiment (dB)

Analysis 1				Configuration	
			Optimal	Current	Gain
SN ratio		nation irmation	12.85 -2.53	-14.01 -17.58	26.87 15.05
Sensitivity	Estimation Confirmation		31.02 30.84	28.62 23.30	2.40 7.54
Analysis 2				Configuration	
			Optimal	Current	Gain
Sn Ratio	<i>T</i> ₁	Estimation Confirmation	29.67 20.01	5.33 7.61	24.34 12.40
	<i>T</i> ₂	Estimation Confirmation	37.77 22.20	5.08 7.88	32.69 14.32
	T ₃	Estimation Confirmation	35.47 23.84	6.54 7.87	29.93 16.44
	T ₅	Estimation Confirmation	40.66 25.65	6.33 7.68	34.33 17.97
	<i>T</i> ₆	Estimation Confirmation	43.20 26.75	6.32 7.69	36.88 19.05
	T ₇	Estimation Confirmation	42.77 28.55	6.40 7.70	36.37 20.85
Sensitivity	<i>T</i> ₁	Estimation Confirmation	-0.19 -0.07	$-0.14 \\ -0.10$	-0.05 0.02
	<i>T</i> ₂	Estimation Confirmation	-0.13 -0.04	$-0.10 \\ -0.08$	-0.03 0.04
	T ₃	Estimation Confirmation	-0.12 -0.03	-0.08 -0.06	-0.04 0.03
	Τ ₄	Estimation Confirmation	-0.10 -0.03	-0.04 -0.05	-0.05 0.02
	T ₅	Estimation Confirmation	-0.08 -0.02	-0.02 -0.04	-0.06 0.02
	T ₆	Estimation Confirmation	-0.07 -0.02	-0.0 -0.02	-0.07 0.01
	<i>T</i> ₇	Estimation Confirmation	-0.06 -0.01	-0.01 -0.02	-0.05 0.01

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Streamlined Inspection due to Time Reduction If we assume that

inspection cost = (sample cost per inspection) + (labor cost) + (machine running cost)

where the daily labor cost is 10,000 yen and the machine running cost is 155 yen, we obtain the following costs:

Current configuration: 235 + (10,000)(5) + (155)(5) = 51,010Optimal configuration: 221 + (10,000)(2) + (155)(2) = 20,531

The cost benefit per inspection is 30,479 yen.

Risk Aversion due to Time Reduction

Now suppose that it takes two days from the point when the production of a product is completed to the point when it starts to be sold at a shop. If the optimally produced product contains a harmful bacterium, despite being in process of shipment, we can withdraw it shortly after checking the result of inspection. Then we assume that the loss in withdrawal costs 1 million yen. On the other hand, under the current configuration, the product reaches a consumer. In this case, we suppose that the loss to a consumer amounts to 150 million yen per person for the death, or 100,000 yen for hospitalization (as the number of victims is derived from a certain past record).

Current:
$$\frac{(180 \text{ persons} \times 100,000 \text{ yen})}{+ (3 \text{ persons} \times 150 \text{ million yen})}$$
$$= 2,557,377 \text{ yen/product}$$
$$Optimal: \frac{1 \text{ million yen}}{183}$$
$$= 5,264 \text{ yen/product}$$

Thanks to time reduction, we can slash the social loss by 2,557,377 - 5264 = 2,552,113 yen/product.

Loss Function of Detectability of Bacteria

Now we assume that if 100,000 harmful bacteria exist in a certain product, a person who eats it will die, and the resulting loss totals 150 million yen (for loss of life). On the other hand, the loss due to discard of a product is 300 yen.

$$A_0 = 150$$
 million yen
 $A = 300$ yen
 $\Delta_0 = 100,000$ products

The tolerance is

$$\Delta = \sqrt{\frac{A}{A_0}} \Delta_0$$
$$= \sqrt{\frac{300}{150,000,000}} (100,000) = 141.42$$

Supposing that the current average number of bacteria is 100, we have the following loss function:

$$L = \frac{300}{(141.42)^2} (100^2) = 150 \text{ yen}$$

Converting the gain of 20.85 dB into an antilog number, we obtain 121.62. Then, the improvement achieved is (150)(1/121.62) = 1.23. Therefore, the current loss is 150 yen, whereas the optimal is 1 yen. As a result, we can reduce the loss by 150 - 1 = 149 yen.

This study took only six days to complete. In our conventional process, we have had an enormous amount of time loss through reiterated trials and errors, and moreover, always doubted whether we could arrive at a conclusion. If research work can be completed within a short period by taking advantage of quality engineering, we believe that many achievements can be obtained for other inspection methods.

Reference

Eiko Mikami and Hiroshi Yano, 2001. Studies on the method for detection of thermoresistant bacteria. *Proceedings of the 9th Quality Engineering Symposium*, pp. 126–129.

This case study is contributed by Eiko Mikami and Hiroshi Yano.